Trypanorhynch cestodes of teleost fish from the Persian Gulf, Iran

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(Accepted 2 August 2010)

Abstract

A total of 299 fish belonging to ten teleost species were studied in Iranian waters at the north-eastern coast of the Persian Gulf for larval trypanorhynch cestode infection. The following trypanorhynch species were identified: Callitetrarhynchus gracilis Pintner, 1931, Pseudogilquinia thomasi (Palm, 2000), Pterobothrium lesteri Campbell and Beveridge, 1996 and Pseudolacistorhynchus shipleyi (Southwell, 1929). The most abundant parasite was C. gracilis which was isolated from seven teleost species. The highest prevalence (62.9%) and dominance (0.98) were demonstrated for *P. thomasi* with a maximum intensity of 22 in Indian halibut Psettodes erumei. Collections of P. lesteri and P. shipleyi from the Persian Gulf represent new locality records, thus extending the known range of distribution for both species. Four new host records are established. The recorded combination of trypanorhynch species and the established host range correspond to other regions, such as off the Brazilian coast. In terms of species distribution, the north-western part of the Indian Ocean, together with the Gulf of Bengal and the Indonesian archipelago, share the same species, resulting from connected water bodies. All trypanorhynch species were isolated from the body cavity, except for P. thomasi which was recorded from the musculature of P. erumei. Relatively high intensities of infection in P. erumei may be of public health concern in the region, necessitating the consumption of well-cooked fish products.

Introduction

Fish parasites can have a negative impact on the commercial fisheries industry (Mehl, 1970; Grabda, 1977; Palm & Overstreet, 2000a). Although only a few cases of accidental human infections by trypanorhynchs have been reported (Palm, 2004), these worms may cause allergic reactions (Pelayo *et al.*, 2009). As a consequence of the highly developed large-scale fishing industry in the northern part of the Persian Gulf (Taghavi Motlagh *et al.*, 2006), thorough studies on the marine fish parasite fauna are of major importance for the region.

Trypanorhynch cestodes mature in elasmobranchs and mainly use teleost fishes as intermediate or paratenic hosts (e.g. Palm, 1997, 2000). Because of the possibility of identifying adults as well as their larval stages based on the oncotaxy of unique hook arrangements along tentacles (Campbell & Beveridge, 1994; Palm, 2004), these marine tapeworms have been considered an interesting group for taxonomic and ecological investigations (Palm & Klimpel, 2007). Of special interest is information concerning the host specificity within the taxon (Palm & Caira, 2008), providing better insight into the zoogeographical species distribution (Jakob & Palm, 2006; Palm *et al.*, 2007, 2009). Extensive fish parasitological surveys on the trypanorhynch cestode fauna in the Indian Ocean have been carried out by Palm (2000, 2004) and Palm & Overstreet (2000b) off the Indonesian coast and by Southwell (1929, 1930) off Ceylon. These authors revealed a species-rich fauna, suggesting that the former region, especially, is one of the centres of trypanorhynch species

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distribution. An observed level of low host specificity (Palm & Caira, 2008) questions how far these species extend into the western or north-western part of the Indian Ocean.

The Persian Gulf is a semi-closed water body that is connected to the Gulf of Oman through the narrow Hormuz Strait. The Gulf of Oman is the most northwesterly part of the Indian Ocean that opens into the Arabian Sea. Studies on the trypanorhynch fauna of fishes from the Persian Gulf are non-comprehensive (Mirzayans, 1970; Tirgari et al., 1975; Ghiasi, 1988; El-Naffar et al., 1992; Saif et al., 1994; Sheini mandani, 1994; Sazandegi, 1996; Kardousha, 1999; Hassan et al., 2002: Peighan et al., 2004: Haseli, 2005), and focused on the most eastern region. Most recently, Haseli et al. (2010) studied elasmobranchs along the Iranian coast of the Persian Gulf and identified 22 taxa. These studies demonstrate that trypanorhynch cestodes are common fish parasites in the Persian Gulf, corresponding with a diverse fish fauna in the region (Blegvad & Lopenthin, 1944; Kuronuma & Abe, 1972; Randall et al., 1978; Fischer & Bianchi, 1984; Kuronuma & Abe, 1986; Assadi & Dehghani, 1997; Carpenter et al., 1997).

The purpose of the present study is an examination of the trypanorhynch fauna of ten commercially important teleosts from the Iranian coast of the Persian Gulf. In combination with a study of trypanorhynch cestodes from elasmobranchs from the same region (Haseli et al., 2010), the present investigation compiles the known trypanorhynch fauna from the region. New host and locality records extend the known range of distribution for trypanorhynchs formerly known from the eastern part of the Indian Ocean into the Persian Gulf. For the first time, the dominance index was used in order to evaluate the infection pattern, and the Sorenson qualitative and quantitative indices to evaluate the similarity of trypanorhynch infections. To date, none of these ecological metrics have been applied to analyse the trypanorhynch infection pattern within teleost intermediate hosts.

Materials and methods

Collection and examination of fish

A total of 299 fish, comprising *Psettodes erumei* (N = 35, Psettodidae); Silver sillago, *Sillago sihama* (N = 35, Sillaginidae); Indian mackerel, *Rastrelliger kanagurta* (N = 35, Scombridae); Tigertooth croaker, *Otolithes ruber*

(N = 35, Sciaenidae); Greater lizardfish, Saurida tumbil (N = 35, Synodontidae); Whitefin wolf-herring, *Chirocentrus nudus* (N = 35, Chirocentridae); John's snapper, Lutianus johnii (N = 35, Lutianidae); Spangled emperor, *Lethrinus nebulosus* (N = 15, Lethrinidae); Yellowfin seabream, Acanthopagrus latus (N = 35, Sparidae) and Hound needlefish, Tylosurus crocodilus crocodilus (N = 4, Belonidae) were bought in the fish market of Bandar Abbas, on the north-eastern coast of the Persian Gulf, Iran (27°11'N, 56°16'E) in April 2008. Identification was carried out according to published keys for the Persian Gulf (Assadi & Dehghani, 1997; Carpenter et al., 1997). Following biometry (table 1), the fish were examined macroscopically with the help of a stereo-microscope under $6.3 \times$ magnification for larval trypanorhynch cestodes in the body cavity, the viscera and the musculature. Encapsulated trypanorhynch larvae were freed from the blastocyst and treated according to Palm (2004). The worms were fixed in 70% ethanol and stained in acetic carmine, dehydrated in an ethanol series, cleared in methyl salicylate and mounted on to slides in Canada balsam. Trypanorhynch species identification followed Palm (2004).

Data analyses

Data analyses of the parasites and hosts were done using SPSS Package, version 13 (SPSS Inc., Chicago, Illinois, USA). According to Magurran (1988), the dominance index of the most dominant parasite species in each host (dominance index = number of individuals of the most abundant species/total number of individuals in the community) and the Sorenson qualitative ($C_{\rm S} =$ twice the number of species found in common/the total number of species that are present in two communities) and quantitative (C_N = twice the sum of the lower of the two abundances for the recorded species/the total number of individuals that are present in two communities) indices were calculated. Unlike the Sorenson qualitative index, which counts all recorded species equally irrespective of whether they are abundant or rare, the Sorenson quantitative index takes into account abundances of respective species. Thus, a better idea of the trypanorhynch cestode infection pattern can be presented by using both similarity indices. Prevalence, mean abundance, intensity and mean intensity were calculated according to Bush et al. (1997). The specificity index (H_S) of Caira et al. (2003) was used to assess host

Table 1. Number, total length and sex of examined teleost species.

Family	Fish species	Number	Total length (cm)	Sex (male: female)
Psettodidae	Psettodes erumei	35	34-44	24:11
Sillaginidae	Sillago sihama	35	16-26	7:28
Scombridae	Rastrelliger kanagurta	35	17.5-22	11:24
Sciaenidae	Otolithes ruber	35	30-41	9:26
Synodontidae	Saurida tumbil	35	24-51	0:35
Ćhirocentridae	Chirocentrus nudus	35	29-50	3:32
Lutjanidae	Lutjanus johnii	35	29-50	18:17
Lethrinidae	Lethrinus nebulosus	15	37.5-54	10:5
Sparidae	Acanthopagrus latus	35	22-34	12:23
Belonidae	Tylosurus crocodilus crocodilus	4	93–111	0:4

Parasite species	Host species	Prevalence (%)	Intensity	Mean intensity (±SE)	Mean abundance (±SE)
Family Lacistorhynchidae Guiart,	1927				
Callitetrarhynchus gracilis	Psettodes erumei	5.7	1	1 ± 0	0.06 ± 0.04
$H_S = 8.2624$	Otolithes ruber	5.7	1	1 ± 0	0.06 ± 0.04
	Saurida tumbil	14.3	1-9	3.6 ± 1.44	0.52 ± 0.28
	Chirocentrus nudus	28.6^{*}	1 - 4	2 ± 0.33	0.57 ± 0.18
	Lutjanus johnii	8.6	1-3	2 ± 0.58	0.17 ± 0.10
	Lethrinus nebulosus	60	1-12	4.22 ± 1.14	2.53 ± 0.87
	Tylosurus crocodilus crocodilus	25*	4	4	1 ± 1
Pseudogilquinia thomasi $H_{\rm S} = 0.00$	Psettodes erumei ⁺	62.9	1–22	5.86 ± 1.20	3.68 ± 0.89
Pseudolacistorhynchus shipleyi $H_{\rm S} = 0.00$	Lethrinus nebulosus	6.7*	1	1	0.07 ± 0.07
Family Pterobothriidae Pintner, 19 <i>Pterobothrium lesteri</i> $H_S = 0.00$	031 Otolithes ruber	25.7 [*]	1–2	1.11 ± 0.11	0.28 ± 0.09

Table 2. Prevalence, intensity, mean intensity, mean abundance and infection site of trypanorhynch cestodes from ten teleost species from the Persian Gulf. New host records are marked by an asterisk after prevalence values. H_S according to Caira *et al.* (2003).

⁺ All trypanorhynch species were isolated from the body cavity, except for *P. thomasi* which was recorded from the musculature of *P. erumei*.

specificity. Voucher specimens have been deposited in the ZUTC, collection of the Zoological Museum, University of Tehran, Tehran, Iran (ZUTC Platy 1223–5, *Callitetrarhynchus gracilis*; ZUTC Platy 1214–7, *Pseudogilquinia thomasi*; ZUTC Platy 1218–22, *Pterobothrium lesteri*; ZUTC Platy 1226, *Pseudolacistorhynchus shipleyi*) and the Museum Berlin, Germany (ZMB).

Results

Identification of cestodes and levels of infection

Four species of trypanorhynch cestodes, *Callitetrarhynchus gracilis, Pseudogilquinia thomasi, Pseudolacistorhynchus shipleyi* and *Pterobothrium lesteri*, were found in 7 out of 10 studied fish species. *Pterobothrium lesteri* and *P. shipleyi* represent new locality records from the Persian Gulf, and four new host records are established (table 2). All trypanorhynch species were isolated from the body cavity, except for *P. thomasi* which was recorded from the musculature of *Psettodes erumei*.

The most common parasite was *C. gracilis*, recovered from seven different teleost species. The highest prevalence (62.9%) was seen for *P. thomasi* with a maximum intensity of 22 in *P. erumei*. The Kolmogorov–Smirnov test demonstrated that the parasite infection was not normally distributed; therefore non-parametric tests, Mann–Whitney test (*U*) and Kruskal–Wallis test, were applied to determine whether significant differences in mean abundances of parasites exist. Prevalence, intensity, mean intensity and mean abundance of the trypanor-hynch cestodes from ten teleost species from the Persian Gulf are given in table 2.

Statistical analyses revealed that the prevalence and mean abundance of *C. gracilis* in *C. nudus* were significantly higher than in *P. erumei* ($\chi^2 = 6.437$, P = 0.011, U = 466.50, P = 0.009), *O. ruber* ($\chi^2 = 6.437$, P = 0.011, U = 466.50, P = 0.09) and Lutjanus johnii ($\chi^2 = 4.629$, P = 0.031, U = 490.50, P = 0.035). Prevalence

and mean abundance of *C. gracilis* in *L. nebulosus* were significantly higher than in *P. erumei* ($\chi^2 = 18.032$, P < 0.005, U = 112, P < 0.005), *O. ruber* ($\chi^2 = 18.032$, P < 0.005, U = 112, P < 0.005), *S. tumbil* ($\chi^2 = 10.884$, P = 0.001, U = 139, P = 0.001), *C. nudus* ($\chi^2 = 4.402$, P = 0.036, U = 159, P = 0.012) and *L. johnii* ($\chi^2 = 15.226$, P < 0.005, U = 121.50, P < 0.005).

Cestode species richness

The species richness and dominance indices are summarized in table 3. Apart from *P. erumei, O. ruber* and *L. nebulosus*, which had a species richness of two parasite species, this value was 1 for other infected hosts. The Sorenson qualitative similarity index showed the highest value (1) for all inter-specific comparisons of hosts that were infested with *C. gracilis*. Values were comparatively intermediate between other infested hosts. Among inter-specific comparisons of hosts in which qualitative

Table 3. Species richness and dominance index of trypanorhynch species in ten teleost hosts.

Host species	Species richness	Dominant parasite species	Dominance index
Psettodes erumei	2	P. thomasi	0.98
Sillago sihama	0	_	0
Rastrelliger kanagurta	0	_	0
Otolithes ruber	2	P. lesteri	0.83
Saurida tumbil	1	C. gracilis	1
Chirocentrus nudus	1	C. gracilis	1
Lutjanus johnii	1	C. gracilis	1
Lethrinus nebulosus	2	C. gracilis	0.97
Acanthopagrus latus	0	_	0
Tylosurus crocodilus crocodilus	1	C. gracilis	1

Host species	Host species	$C_{\rm S}$	$C_{\rm N}$	Host species	Host species	$C_{\rm S}$	$C_{\rm N}$
Psettodes erumei	Sillago sihama	0	0	Rastrelliger kanagurta	Tylosurus crocodilus crocodilus	0	0
Psettodes erumei	Rastrelliger kanagurta	0	0	Otolithes ruber	Šaurida tumbil	0.67	0.13
Psettodes erumei	Otolithes ruber	0.5	0.03	Otolithes ruber	Chirocentrus nudus	0.67	0.13
Psettodes erumei	Saurida tumbil	0.67	0.03	Otolithes ruber	Lutjanus johnii	0.67	0.22
Psettodes erumei	Chirocentrus nudus	0.67	0.03	Otolithes ruber	Lethrinus nebulosus	0.5	0.08
Psettodes erumei	Lutjanus johnii	0.67	0.03	Otolithes ruber	Acanthopagrus latus	0	0
Psettodes erumei	Lethrinus nebulosus	0.5	0.02	Otolithes ruber	Tylosurus crocodilus crocodilus	0.67	0.25
Psettodes erumei	Acanthopagrus latus	0	0	Saurida tumbil	Čhirocentrus nudus	1	0.95
Psettodes erumei	Tylosurus crocodilus crocodilus	0.67	0.03	Saurida tumbil	Lutjanus johnii	1	0.5
Sillago sihama	Rastrelliger kanagurta	0	0	Saurida tumbil	Lethrinus nebulosus	0.67	0.92
Sillago sihama	Otolithes ruber	0	0	Saurida tumbil	Acanthopagrus latus	0	0
Sillago sihama	Saurida tumbil	0	0	Saurida tumbil	Tylosurus crocodilus crocodilus	1	0.36
Sillago sihama	Chirocentrus nudus	0	0	Chirocentrus nudus	Lutjanus johnii	1	0.46
Sillago sihama	Lutjanus johnii	0	0	Chirocentrus nudus	Lethrinus nebulosus	0.67	0.68
Sillago sihama	Lethrinus nebulosus	0	0	Chirocentrus nudus	Acanthopagrus latus	0	0
Sillago sihama	Acanthopagrus latus	0	0	Chirocentrus nudus	Tylosurus crocodilus crocodilus	1	0.33
Sillago sihama	Tylosurus crocodilus crocodilus	0	0	Lutjanus johnii	Lethrinus nebulosus	0.67	0.27
Rastrelliger kanagurta	<i>Ótolithes ruber</i>	0	0	Lutjanus johnii	Acanthopagrus latus	0	0
Rastrelliger kanagurta	Saurida tumbil	0	0	Lutjanus johnii	Tylosurus crocodilus crocodilus	1	0.8
Rastrelliger kanagurta	Chirocentrus nudus	0	0	Lethrinus nebulosus	Ăcanthopagrus latus	0	0
Rastrelliger kanagurta	Lutjanus johnii	0	0	Lethrinus nebulosus	Tylosurus crocodilus crocodilus	0.67	0.19
Rastrelliger kanagurta	Lethrinus nebulosus	0	0	Acanthopagrus latus	Tylosurus crocodilus crocodilus	0	0
Rastrelliger kanagurta	Acanthopagrus latus	0	0	, 5	<i>.</i>		

Table 4. Sorenson qualitative similarity index (C_S) and Sorenson quantitative similarity index (C_N) between helminth communities of different teleost species.

similarity indices were the highest (1), interestingly, high, low and intermediate values of Sorenson quantitative similarity were observed. Low values of Sorenson quantitative similarity were mostly calculated for interspecific comparisons of hosts with intermediate values of Sorenson qualitative similarity (table 4).

Discussion

The present study provides the first information on a spectrum of trypanorhynch infections of ten teleost species from the Iranian waters of the north-eastern Persian Gulf. Until now the larval trypanorhynch fauna had been studied mainly from the southern part of the Persian Gulf (El-Naffar *et al.*, 1992; Saif *et al.*, 1994; Kardousha, 1999; Hassan *et al.*, 2002). Summaries of reported larval and adult trypanorhynchs from the

Persian Gulf are given in tables 5–7 (see also Palm, 2004; Haseli *et al.*, 2010).

Of the recorded four trypanorhynch species, three species of the superfamily Lacistorhynchoidea Guiart, 1927, namely, P. lesteri, P. thomasi and P. shipleyi, demonstrated restricted host range. They were found in O. ruber, P. erumei and L. nebulosus, respectively, indicating higher specificity within the sampled subset of predatory fish. This is in contrast to the occurrence of the cosmopolitan C. gracilis (see Palm, 2004) which infected 7 of 10 fish species. This situation is similar to an earlier investigation along the north-east Brazilian coast by Palm (1997). Within that study, C. gracilis was the trypanorhynch cestode with the widest host range, which could be isolated from 10 fish species out of the 57 sampled fish species from 30 families. The co-infection with another pseudolacistorhynchid. Pseudolacistorhynchus noodti, from Pseudupeneus maculatus and Scomberomorus maculatus, and

Table 5. Reported larval trypanorhynchs belonging to families Tentaculariidae, Rhopalothylacidae and Pterobothriidae in teleost species from the Persian Gulf.

Parasite species	Fish host	Location
Family Tentaculariidae		
<i>Tetrarhynchus</i> sp. ⁷	Epinephelus coioides	Ι
Tentacularia coryphaenae ²	Euthynnus affinis	A,B,D,G
Nybelinia sp. ¹	Saurida tumbil	В
J 1	Upeneus tragula	А
Nybelinia indica ^{2&6}	Alepes djedaba ²	G
5	Epinephelus tauvina ⁶	Н
	Nemipterus Japonicus ⁶	Н
	Upeneus tragula ⁶	Н
Heteronybelinia elongata ^{1&2}	Lethrinus lentjan ²	G
3	Lutjanus kasmira ²	G
	Pampus argenteus ^{1&2}	B,G
	Saurida tumbil ²	G
<i>Heteronybelinia</i> sp. ^{1&2}	Trichiurus lepturus ^{1&2}	D,G
Family Rhopalothylacidae	1	,
Pintneriella musculicola ^{4&6}	Epinephelus chlorostigma ⁶	Н
Pinineriella musculicola	Epinephelus tauvina ⁴	E
	Lethrinus nebulosus ⁶	H
Family Pterobothriidae		
Pterobothrium sp. ^{1,2,5&9}	Aesopia cornuta ¹	D
<i>i icrobolinium s</i> p.	Caranx sp. ⁵	C
	Cephalopholis miniata ²	G
	Epinephelus areolatus ¹	B
	Epinephelus chlorostigma ¹	A
	Saurida tumbil ¹	A
	Sphyraena jello ¹	D
	Thunnus tonggol ⁹	C
	Trichiurus lepturus ¹	A
Pterobothrium heteracanthum ²	Aesopia cornuta	G
1 terobotinitumi neterucuntitumi	Arius thalassinus	G
	Carangoides malabaricus	G
	Epinephelus areolatus	G
	Epinephelus chlorostigma	G
	Psettodes erumei	G
	Saurida tumbil	G
		G
	Sphyraena jello Trichiurus Ienturus	G
	Trichiurus lepturus	G

^{1.} El-Naffar *et al.* (1992); 2. Kardousha (1999); 3. Haseli (2005); 4. Hassan *et al.* (2002); 5. Mirzayans (1970); 6. Palm (2004); 7. Peighan *et al.* (2004); 8. Saif *et al.* (1994); 9. Sazandegi (1996); 10. Tirgari *et al.* (1975). A. Abu Dhabi; B. Dubai and Sharjah; C. Iranian waters; D. Ras al Khaimah; E. The coasts of Bahrain, Oman, Qatar and the United Arab Emirates; F. The coast of Qatar; G. The coast of UAE; H. The Persian Gulf; I. Western part of the Persian Gulf, Iran.

Table 6. Reported larval trypanorhynchs belonging to families Lacistorhynchidae, Pseudotobothriidae and Otobothriidae in teleost species from the Persian Gulf. See table 5 for abbreviations.

Parasite species	Fish host	Location	Parasite species	Fish host	Location
Family Lacistorhynchidae			Family Lacistorhynchidae		
Protogrillotia arabiensis ⁶	Siganus javus	Н	Floriceps sp. ^{1&6}	Aesopia cornuta	А
Pseudogrillotia sp. ²	Lethrinus erythracanthus	G		Alepes djedaba	А
Pseudogrillotia spratti ²	Liza macrolepis	G		Carangoides malabaricus	А
Pseudogilquinia thomasi ⁶	Psettodes erumei	Ĥ		Cephalopholis miniata	В
Pseudogilquinia kardoushi ⁶	Argyrops filamentosus	H		Epinephelus areolatus	B
i bennoginquinna naraoubin	Lethrinus erythracanthus	H		Epinephelus chlorostigma	B
	Psettodes erumei	H		Epinephelus tauvina	Ă
Pseudogilquinia microbothria ⁶	Parupeneus ciliatus	H		Lethrinus erythracanthus	D
Grillotia sp. ¹⁰	Thunnus thynnus	C		Lethrinus lentjan	D
<i>Callitetrarhynchus gracilis</i> ^{2&6}	Alepes djedaba ²	G		Lethrinus nebulosus	A
Cutilietramynenus graenis	Euthynnus affinis ^{2&6}	G		Liza macrolepis	A
	Lethrinus erythracanthus ²	G		Lutjanus coccineus	В
	Nemipterus japonicus ²	G		Lutjanus tulviflamma	B
	Saurida tumbil ²	G		Lutjanus kasmira	D
	Scomberoides commersonnianus ²	G			B
		-		Nemipterus peronii	
	Sphyraena barracuda ²	G,H		Pampus argenteus	D
C 11:1 1 1 : 2.4&6	Trichiurus lepturus ²	G		Parastromateus niger	В
<i>Callitetrarhynchus speciosus</i> ^{2,4&6}	Arius thalassinus ⁶	H		Pomadasys argenteus	D
	<i>Cephalopholis hemistiktos</i> ⁶	Н		Saurida undosquamis	D
	Epinephelus areolatus ²	G		Sphyraena jello	В
	Lethrinus nebulosus ⁴	E		Upeneus tragula	A
<i>Callitetrarhynchus</i> sp. ^{2&7}	Argyrops filamentosus ²	G	Floriceps minacanthus ⁶	Cephalopholis hemistiktos	Η
	Carangoides malabaricus ²	G	Family Pseudotobothriidae		
	Cephalopholis miniata ²	G	Pseudotobothrium dipsacum ^{6&2}	Epinephelus chlorostigma ⁶	F
	Epinephelus chlorost <u>i</u> gma ²	G		Epinephelus tauvina ⁶	F
	Epinephelus coioides ⁷	Ι		Psettodes erumei ²	Н
	Epinephelus tauvina ²	G	Family Otobothriidae		
	Lethrinus nebulosus ²	G	Symbothriorhynchus tigaminacantha ⁶	Nemipterus japonicus	Н
	Lutjanus coccineus ²	G		, , ,	
	Lutjanus fulviflamma ²	G			
	Lutjanus johnii ²	G			
	Lutjanus kasmira ²	G			
	Nemipterus peronii ²	G			
	Psettodes erumei ²	G			
	Saurida undosquamis ²	Ğ			
	Selaroides leptolepis ²	G			
	Sphyraena jello ²	G			

Table 7. Reported	adult trypanorhynchs	from the Persian Gulf.

Parasite species	Fish host	Parasite species	Fish host
Family Tentaculariidae		Family Eutetrarhynchidae	
Kotorella pronosoma	Himantura cf. uarnak	Eutetrarhynchus sp.	Himantura imbricata
,	Rhynchobatus cf. djiddensis	v I	Rhinobatos cf. punctifer
Kotorella sp.	Himantura imbricata	Oncomegoides celatus	Himantura sp.
Nybelinia sp. I	Rhizoprionodon acutus	Parachristianella monomegacantha	Himantura cf. uarnak
Nybelinia sp. II	Himantura imbricata	0	Pastinachus cf. sephen
Heteronybelinia heteromorphi	Carcharhinus cf. dussumieri		Rhynchobatus cf. djiddensis
Family Pterobothriidae		Parachristianella indonesiensis	Himantura cf. uarnak
Pterobothrium lesteri	Gymnura cf. poecilura		Himantura sp.
Family Lacistorhynchidae	eginnin en pecennin		Pastinachus cf. sephen
Pseudogrillotia perelica	Carcharhinus cf. dussumieri		Rhynchobatus cf. djiddensis
Pseudogrillotia sp.	Carcharhinus cf. dussumieri	Parachristianella sp.	Himantura imbricata
Callitetrarhynchus gracilis	Carcharhinus cf. dussumieri	Pseudochristianella southwelli	Rhinobatos cf. punctifer
Cuttitetrumynenus gruettis	Carcharhinus cf. sorrah	Prochristianella macracantha	Pastinachus cf. sephen
	Rhizoprionodon acutus	Prochristianella spp.	Aetomylaeus cf. nichofii
Family Otobothriidae	Knizoprionouon ucuius	Froenrisiuneitu spp.	Himantura imbricata
Proemotobothrium southwelli	Carcharhinus cf. dussumieri		Himantura sp.
Froemotobolinnum Southwelli			
	<i>Gymnura</i> cf. poecilura	Dellfusielle	Pastinachus cf. sephen
Due and take the internet and	Rhizoprionodon acutus Carcharhinus cf. dussumieri	Dollfusiella spp.	Aetomylaeus cf. nichofii
Proemotobothrium sp. Otobothrium carcharidis			Aetomylaeus maculatus
Otobothrium carcharlais	Carcharhinus cf. dussumieri		Himantura cf. uarnak
	Carcharhinus cf. sorrah		Himantura imbricata
	Rhizoprionodon acutus		<i>Himantura</i> sp.
<i>Otobothrium</i> sp.	Carcharhinus cf. dussumieri		Pastinachus cf. sephen
	Rhizoprionodon acutus		Rhinobatos cf. punctifer
Family Mixodigmatidae			
Halysiorhynchus macrocephalus	Pastinachus cf. sephen		
	Rhynchobatus cf. djiddensis		
Trygonicola macropora	<i>Himantura</i> sp.		
	Himantura cf. uarnak		
	Pastinachus cf. sephen		
	Rhinobatos cf. punctifer		
Family Eutetrarhynchidae			
Eutetrarhynchus platycephali	<i>Himantura</i> sp.		
	Pastinachus cf. sephen		
	Rhinobatos cf. punctifer		

another pterobothriid *Pterobothrium* sp. from *Citharichthys spilopterus*, showed a similar narrow host range as was observed within the present study. It seems as if the recorded combination of representatives from these three lacistorhynchoid families and the established host range are similar along the tropical north-eastern coast of Brazil and the Persian Gulf.

The record of C. gracilis from a range of different hosts in the Persian Gulf supports earlier investigations (e.g. Palm, 1997), and the low host specificity (Palm & Caira, 2008) and cosmopolitan distribution (Palm, 2004) of the species. Similarly, Abdou & Palm (2008) recorded C. gracilis in a range of different intermediate hosts in the Red Sea. Such a distribution pattern is promoted by the omnipresence of a variety of carcharhinid elasmobranchs as definitive hosts in coastal tropical regions, combined with restricted host specificity in both second intermediate and definitive hosts. Palm et al. (1994) and Palm (1997) proposed a four-host cycle for C. gracilis, including small pelagic Carangidae and Clupeidae for the first stage, and larger predatory fish, mostly Carangidae and Scombridae, for the second stage of the plerocerci. Palm & Caira (2008) showed that the average value of host specificity

 $(H_{\rm S})$ for final larval stages of 63 trypanorhynch species was 6.29 (euryxenous), and the value for *C. gracilis* was 8.61 (euryxenous). The $H_{\rm S}$ recorded herewith for *C. gracilis* from teleosts in the Persian Gulf was 8.26, also euryxenous. Two new host records from *C. nudus* and *T. crocodilus* crocodilus also suggest that more studies can further extend the already known wide host range for *C. gracilis*.

The trypanorhynch cestode fauna recorded so far from the Persian Gulf demonstrates a predominance of eutetrarhynchoid and lacistorhynchoid species, with only few tentacularioids or gymnorhynchoids. This might be explained with missing faunistic components in the Gulf, such as big pelagic sharks that are common hosts for gymnorhynchoids. Another possible explanation is the lack of potential first intermediate hosts, the most important stage in the completion of the life cycle of trypanorhynchs (Palm *et al.*, 1994). Although El-Naffar *et al.* (1992) and Kardousha (1999) reported some tentaculariid species from the south of the Persian Gulf, not a single specimen belonging to the superfamily Tentacularioidea Poche, 1926 has been reported from the sampled teleosts in the northern part of the Persian Gulf.

According to Palm (2004), the life cycle includes four or even more hosts, with copepods as first, euphausiids or schooling fish as second intermediate hosts and fish as third or more intermediate or paratenic hosts. Food resources, depth, salinity and temperature might influence the distribution and species composition of required copepod first intermediate hosts (Islam et al., 2005; Gaard et al., 2008). Euphausiids as second intermediate hosts for tentaculariids might also be unevenly distributed. Zhaoli (2007) and Yoon et al. (2000) demonstrated that temperature, salinity and chlorophyll a concentration are important ecological factors that determine spatial distribution of euphausiids. Hunter (1983) and Revnolds (1993) stated that the salinity of the counterclockwise water current in the Persian Gulf can be influenced by rivers in the north (e.g. Tigris, Euphrates) leading to less saline waters in the Strait of Hormuz at the surface. Emery (1956) demonstrated that the bottom of the Persian Gulf is mostly muddy in the north and mostly sandy in the south, and Ebrahimi et al. (2008) showed that salinity increased horizontally toward the west of the Persian Gulf. These ecological differences in the north and south of the Persian Gulf might play a role in the uneven distribution of the recorded trypanorhynchs.

The highest value (1) of the Sorenson qualitative similarity and low values of Sorenson quantitative similarity indices can be explained by the low diversity and high dominance of the trypanorhynch component community in the sampled fish. The high Sorenson qualitative similarity (1) combined with the high quantitative similarity (0.95) that was observed for *Š. tumbil* and *C. nudus* can be explained by a high prevalence and mean abundance of *C. gracilis* in both hosts. It is worth mentioning that there was no significant difference of the prevalence and mean abundance in these hosts, though representing different fish families. This can only be explained by a similar size and feeding ecology of these species (Fischer et al., 1990; Sommer et al., 1996), demonstrating that the feeding ecology of the fish must have a major important role for infection with C. gracilis in the Persian Gulf.

Regarding the high diversity of trypanorhynchs in the Persian Gulf (Haseli *et al.*, 2010), this semi-enclosed marine ecosystem has the capacity for the completion of a variety of life cycles. According to Palm (2004) and Southwell (1929, 1930), it seems that the east of the Indian Ocean, the Indonesian archipelago and Sri Lanka share the same species, resulting from connected water bodies. The Persian Gulf can be invaded by trypanorhynchs due to their low host specificity (Palm & Caira, 2008) and wide zoogeographical distribution of many of their host fish species (Fischer & Whitehead, 1974; Compagno, 1984; Fischer & Bianchi, 1984; Carpenter *et al.*, 1997).

Among the ten studied commercially important teleosts in the Iranian waters of the Persian Gulf, musculature infection was restricted to *P. erumei* at a prevalence of 62.9% and a range of intensity of 1–22. This relatively high infection of muscles with the large plerocerci of *P. thomasi* may cause concern for marketing and public health (see Pelayo *et al.*, 2009). It is recommended that *P. erumei* be well cooked and freed from the most obvious trypanorhynch larvae before the fish is sold and considered for human consumption.

Acknowledgements

We wish to thank Professor Dr H. Mehlhorn who facilitated the six-month stay of M.H. at Heinrich-Heine-University, Düsseldorf. Financial support was provided through the German Research Council (DFG PA 664/4-1 and 4-2) (H.W.P.) and the Iran Ministry of Science, Research and Technology together with the Centre of Research Affairs of the University of Tehran (M.H.). This study has been carried out within the framework of the NSF Planetary Biodiversity and Inventory Project (PB&I) awards Nos. 0818696 and 0818823.

References

- Abdou, N.E. & Palm, H.W. (2008) New records of trypanorhynch cestodes from Red Sea fishes in Egypt. *Journal of the Egyptian Society of Parasitology* 38, 281–292.
- **Assadi, H. & Dehghani, R.** (1997) Atlas of the Persian Gulf and the Sea of Oman fishes. 226 pp. Tehran, Iranian Fisheries Research and Training Organization.
- **Blegvad, H. & Lopenthin, B.** (1944) *Fishes of the Iranian Gulf.* 247 pp. Copenhagen, Einar Munksgaard.
- Bush, A.O., Lafferty, K.D., Lotz, J.M. & Shostak, A.W. (1997) Parasitology meets ecology on its own terms: Margolis *et al.* revisited. *Journal of Parasitology* 83, 575–583.
- Caira, J.N., Jensen, K. & Holsinger, K. (2003) On a new index of host specificity. pp. 161–201 in Combes, C. & Jordan, J. (Eds) Taxonomy, ecology, and evolution of metazoan parasites. Vol. I. (Livre hommage à Louis Euzet). Perpignan, PUP Presses.
- Campbell, R.A. & Beveridge, I. (1994) Order Trypanorhyncha Diesing, 1863. pp. 51–148 in Khalil, L.F., Jones, A. & Bray, R.A. (Eds) Keys to the cestode parasites of vertebrates. Wallingford, CAB International.
- Carpenter, K.E., Krupp, F., Jones, D.A. & Zajonz, U. (1997) FAO species identification field guide for fishery purposes. Living marine resources of Kuwait, eastern Saudi Arabia, Bahrain, Qatar, and the United Arab Emirates. 293 pp. Rome, FAO.
- **Compagno, L.J.V.** (1984) *Sharks of the world.* 655 pp. Fisheries Synopsis No. 125, Food and Agriculture Organization of the United Nations. Rome, FAO.
- Ebrahimi, M., Mohebi Nouzar, L., Ajlali Khaneghah, K. & Sanjani, M.S. (2008) Spatial and temporal variation of salinity, water density and temperature of sea's waters in Hormozgan Province, Hormoz strait and the Persian Gulf. *Iranian Scientific Fisheries Journal* **17**, 1–14 (in Persian).
- El-Naffar, M.K.I., Gobashy, A., El-Etreby, S.G. & Kardousha, M.M. (1992) General survey of helminth parasite genera of Arabian Gulf Fishes (coast of United Arab Emirates). *Arab Gulf Journal of Scientific Research* 10, 99–110.
- **Emery, K.O.** (1956) Sediments and water of Persian Gulf. Bulletin of the American Association of Petroleum Geologists **40**, 2354–2383.
- Fischer, W. & Bianchi, G. (Eds) (1984) FAO species identification sheets for fisheries purposes, Western Indian Ocean. Vols I–V. Rome, FAO.

- Fischer, W. & Whitehead, P.J.P. (Eds) (1974) FAO species identification sheets for fishery purposes. Eastern Indian Ocean (fishing area 57) and Western Central Pacific (fishing area 71). Vols 1–4. Rome, FAO.
- Fischer, W., Sousa, I., Silva, C., de Freitas, A., Poutiers, J.M., Schneider, W., Borges, T.C., Feral, J.P. & Massinga, A. (1990) Fichas FAO de identificação de espécies para actividades de pesca. Guia de campo das espécies comerciais marinhas e de águas salobras de Moçambique. 424 pp. Rome, FAO.
- Gaard, E., Gislason, A., Falkenhaug, T., Soiland, H., Musaeva, E., Vereshchaka, A. & Vinogradov, G. (2008) Horizontal and vertical copepod distribution and abundance on the Mid-Atlantic Ridge in June 2004. Deep-Sea Research II 55, 59–71.
- **Ghiasi, N.** (1988) The study of the fish and fisheries situation in Bandar Lengeh, how the trypanorhynch larvae infest the fish. Unpublished MSc thesis, University of Tehran, Iran.
- Grabda, J. (1977) Studies on parasitation and consumability of Alaska pollack, *Theragra chalcogramma* (Pall.). *Acta Ichthyologica et Piscatoria* 7, 15–34.
- Haseli, M. (2005) Ecology of tapeworms in the shark, Carcharhinus dussumieri from the Persian Gulf. Unpublished MSc thesis, University of Tehran, Iran.
- Haseli, M., Malek, M. & Palm, H.W. (2010) Trypanorhynch cestodes of elasmobranchs from the Persian Gulf. Zootaxa 2492, 28–48.
- Hassan, M.A., Palm, H.W., Mahmoud, M.A. & Jama, F.A. (2002) Trypanorhynch cestodes from the musculature of commercial fishes from the Arabian Gulf. *Arab Gulf Journal of Scientific Research* 20, 74–86.
- Hunter, J.R. (1983) Aspects of the dynamics of the residual circulation of the Arabian Gulf. pp. 31–42 *in* Gade, H.G., Edwards, A. & Svendsen, H. (*Eds*) *Coastal oceanography*. New York, Plenum Press.
- Islam, M.D.S., Ueda, H. & Tanaka, M. (2005) Spatial distribution and trophic ecology of dominant copepods associated with turbidity maximum along the salinity gradient in a highly embayed estuarine system in Ariake Sea, Japan. *Journal of Experimental Marine Biology and Ecology* **316**, 101–115.
- Jakob, E. & Palm, H.W. (2006) Parasites of commercially important fish species from the southern Java coast, Indonesia, including the distribution pattern of trypanorhynch cestodes. *Verhandlungen der Gesellschaft* für Ichthyologie 5, 165–191.
- Kardousha, M.M. (1999) Helminth parasite larvae collected from Arabian Gulf fish II. First record of some trypanorhynch cestodes from economically important fishes. *Arab Gulf Journal of Scientific Research* 17, 255–276.
- Kuronuma, K. & Abe, Y. (1972) *Fishes of Kuwait*. 123 pp. Kuwait, Kuwait Institute for Scientific Research.
- Kuronuma, K. & Abe, Y. (1986) Fishes of the Arabian (Persian) Gulf. 356 pp. Kuwait, Kuwait Institute for Scientific Research.
- Magurran, A.E. (1988) Ecological diversity and its measurement. 179 pp. London, Chapman and Hall.
- Mehl, J.A.P. (1970) Two flesh parasites of barracouta (Teleosti: Gempylidae) from eastern Cook Strait. *New Zealand Journal of Marine and Freshwater Research* 3, 241–247.

- **Mirzayans, A.** (1970) A case report of a cestoda (*Pterobothrium*) found in fish from Persian Gulf. *Journal of the Veterinary Faculty, University of Tehran, Iran* **26**, 43–48 (in Persian).
- Palm, H.W. (1997) Trypanorhynch cestodes of commercial fishes from northeast Brazilian coastal waters. *Memorias do Instituto Oswaldo Cruz* 92, 69–79.
- Palm, H.W. (2000) Trypanorhynch cestodes from Indonesian coastal waters (East Indian Ocean). Folia Parasitologica 47, 123–134.
- Palm, H.W. (2004) The Trypanorhyncha Diesing, 1863. 710 pp. Bogor, IPB-PKSPL Press.
- Palm, H.W. & Caira, J.N. (2008) Host specificity of adult versus larval cestodes of the elasmobranch tapeworm order Trypanorhyncha. *International Journal for Parasitology* 38, 381–388.
- Palm, H.W. & Klimpel, S. (2007) Evolution of the parasitic life in the ocean. *Trends in Parasitology* 23, 10–12.
- Palm, H.W. & Overstreet, R. (2000a) Otobothrium cysticum (Cestoda: Trypanorhyncha) from the muscle of butterfishes (Stromateidae). Parasitology Research 86, 41–53.
- Palm, H.W. & Overstreet, R. (2000b) New records of trypanorhynch cestodes from the Gulf of Mexico, including Kotorella pronosoma (Stossich, 1901) and Heteronybelinia palliata (Linton, 1924) comb. n. Folia Parasitologica 47, 293–302.
- Palm, H.W., Obiekezie, A.I. & Möller, H. (1994) Trypanorhynch cestodes of commercial inshore fishes of the West African coast. *Aquatic Living Resources* 7, 153–164.
- Palm, H.W., Waeschenbach, A. & Littlewood, D.T.J. (2007) Genetic diversity in the trypanorhynch cestode *Tentacularia coryphaenae* Bosc, 1797: evidence for a cosmopolitan distribution and low host specificity in the teleost intermediate host. *Parasitology Research* 101, 153–159.
- Palm, H.W., Waeschenbach, A., Olson, P. & Littlewood, D.T.J. (2009) Molecular phylogeny and evolution of the Trypanorhyncha Diesing, 1863. *Molecular Phylogenetics and Evolution* 52, 351–367.
- Peighan, R., Hoghoghi Rad, N. & Yousef Dezfouli, A. (2004) Determination of parasitic helminthes in Persian Gulf grouper, (*Epinephelus coioides*), and silver pomfret, (*Stromateus cinereus*). *Pajouhesh and Sazandegi* 62, 49–55 (in Persian).
- Pelayo, V., García-Hernández, P., Puente, P., Rodero, M. & Cuéllar, C. (2009) Seroprevalence of anti-Gymnorhynchus gigas (Trypanorhyncha, Gymnorhynchidae) antibodies in a Spanish population. Journal of Parasitology 95, 778–780.
- Randall, J.E., Allen, G.R. & Smith-Vaniz, W.F. (1978) Illustrated identification guide to commercial fishes. Regional Fishery Survey and Development Project (Bahrain, Iran, Iraq, Kuwait, Oman, Qatar, Saudi Arabia, United Arab Emirates). F1:DP/RAB/71–278/3. 221 pp. Rome, Food and Agriculture Organization of the United Nations, United Nations Development Programme.
- **Reynolds, R.M.** (1993) Physical oceanography of the Gulf, Strait of Hormuz, and the Gulf of Oman: results from the Mt. Mitchell Expedition. *Marine Pollution Bulletin* **27**, 35–60.

- Saif, M., Al-Ghais, S.M. & Kardousha, M.M. (1994) Study on some helminth parasites larvae common in Arabian Gulf fish: a comparison between west and east coasts of U.A.E. Arab Gulf Journal of Scientific Research 12, 559–571.
- Sazandegi, J.E. (1996) Study on parasites in *Thunnus* tonggol. Iranian Fisheries Research and Training Organization, IFRO, Newsletter 6.
- Sheini mandani, P. (1994) The survey of the muscular parasites of *Psettodes erumei*. Unpublished MSc thesis, University of Tehran, Iran.
- Sommer, C., Schneider, W. & Poutiers, J.-M. (1996) FAO species identification field guide for fishery purposes. The living marine resources of Somalia. 376 pp. Rome, FAO.
- **Southwell, T.** (1929) A monograph on cestodes of the order Trypanorhyncha from Ceylon and India, Part 1. *Ceylon Journal of Science, Section B* **15**, 169–317.

- **Southwell, T.** (1930) *The fauna of British India, including Ceylon and Burma. Cestoda.* Vol. I, 391 pp. London, Taylor and Francis.
- Taghavi Motlagh, S.A.A., Akhoundi, M. & Shiri, A.R. (2006) Fishing trend and fisheries potential analyses for the Persian Gulf and Sea of Oman, 1973–2003. *Iranian Scientific Fisheries Journal* 15, 35–44 (in Persian).
- Tirgari, M., Radhakrishnan, C.V. & Howard, B.R. (1975) Occurrence of infection by the cestodes *Grillotia* in Persian Gulf Fish. *American Journal of Veterinary Research* **36**, 703.
- Yoon, W.D., Cho, S.H., Lim, D., Choi, Y.K. & Lee, Y. (2000) Spatial distribution of *Euphausia pacifica* (Euphausiacea: Crustacea) in the Yellow Sea. *Journal of Plankton Research* 22, 939–949.
- Zhaoli, X. (2007) Distribution patterns of pelagic euphausiids in the East China Sea. Acta Ecologica Sinica 27, 3678–3686.